

results are summarized in Table 1. The time curves gave plateaus and the reaction showed an absolute RNA dependence and was RNase sensitive. The reaction product is stable up to 30 min at 37° in 0.1 M triethyl amine-acetic acid buffer, pH 11, while methionyl-RNA during this condition has a half life of about 30 sec.

In vivo incorporations of methyl labelled methionine into both log phase and methionine starved W6, have shown an extensive labelling of the sRNA peak and a smaller but significant incorporation also into rRNA.

Counter-current distribution of sRNA from strain 30S₀A5 gives a fractionation with respect to ability to accept methyl groups. Phase systems D and F⁸ were used in 19-transfer experiments. When assayed with the yeast enzyme, the curve for the methylation was almost the same as the one for the distribution of total sRNA in the bottom phase. — Together our results indicate that the yeast and the *E. coli* enzymes have different specificities. They show also that the methylation reaction is highly specific and that the periodate oxidation and the methionine starvation independently make more sites available for methylation. A full account of this work is in preparation and will be published elsewhere.

The work has been supported by U.S. Public Health Service, grant GM 07576—03. K.G.E. holds a *Sven and Lilly Lawski fellowship*.

Two Fluorescent Amino Acids which Function as Cross-linkages between the Peptide Chains in Resilin, a Rubber-like Protein

Svend Olav Andersen

Zoophysiological Laboratory B, Juliane Mariesvej 36, University of Copenhagen, Denmark

Resilin is a structural protein which occurs in insects. Its most characteristic property is that it behaves as a perfect rubber when swollen with water, and that it is completely insoluble in all solvents which do not break peptide linkages. Its properties are best explained by the assumption that the protein contains some sort of very stable linkages between the peptide chains connecting these to a huge three-dimensional network¹⁻³. These linkages must be of a hitherto unknown nature.

During our investigations of resilin obtained from the desert locust *Schistocerca gregaria* (*Forsskål*) we have isolated two fluorescent compounds⁴ which apparently do not occur in other proteins. Both compounds are phenolic amino acids, and one of them is a diamino dicarboxylic acid and the other is a triamino tricarboxylic acid. If both the amino groups and the carboxylic groups are connected to the other amino acids in the protein by means of normal peptide linkages these compounds can function as cross-linking agents. It has been shown that none of the amino groups are free to react with dinitrofluorobenzene in the native protein, indicating that they are involved in some sort of chemical linkage, but it has not been possible to show whether the carboxylic groups are free or not.

From the amounts of these compounds present in resilin it is possible to calculate the average molecular weight of the peptide chain between two neighbouring cross-linkages, when it is assumed that one of these compounds connects two peptide chains and the other one connects three peptide chains together in one junction point. The average molecular chain weight is then found to be 3000 g. This result is in good agreement with the results obtained by physical measurements made on resilin from dragonflies (*Aeshna* species)³.

1. Berg, P. *J. Biol. Chem.* **222** (1956) 1025.
2. Mandel, L. R. and Borek, E. *Biochem. Biophys. Res. Commun.* **6** (1961) 138.
3. Fleissner, E. and Borek, E. *Proc. Natl. Acad. Sci. U.S.A.* **48** (1962) 1199.
4. Boman, H. G., Boman, I. A. and Maas, W. K. in *Biol. Structure and Function*, I (Eds. T. W. Goodwin and O. Lindberg) p. 297. Academic Press, London 1961.
5. Kurland, C. G. and Maaøe, O. *J. Mol. Biol.* **4** (1962) 193.
6. Boman, H. G. and Hjertén, S. *Arch. Biochem. Biophys. Suppl.* **1** (1962) 276.
7. Zamecnik, P. C., Stephenson, M. L. and Scott, J. F. *Proc. Natl. Acad. Sci. U.S.A.* **46** (1960) 811.
8. Wiesmeyer, H., Kjellin, K. and Boman, H. G. *Biochim. Biophys. Acta* **61** (1962) 625.

1. Weis-Fogh, T. *J. Exptl. Biol.* **37** (1960) 889.
2. Weis-Fogh, T. *J. Mol. Biol.* **3** (1961) 520.
3. Weis-Fogh, T. *J. Mol. Biol.* **3** (1961) 648.
4. Andersen, S. O. *Biochim. Biophys. Acta.* **69** (1963) 249.